

# Progressive Education Society's

# Modern College of Arts, Science and Commerce (Autonomous)

# Ganeshkhind, Pune 411016

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# 2022-23

S.Y. B. Sc. Microbiology

Structure of the B. Sc. course Choice Based Credit System

S.Y.B.Sc	. Microbiology
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Semester	Code	Paper	Paper title	Credit
	23-MB-231	Ι	Medical Microbiology and Immunology	2
III	23-MB-232	II	Bacterial Physiology and Fermentation Technology	2
	23-MB-233	III	Practical Course based on theory papers 23-MB-231 and 23-MB-232	2
IV	23-MB-241	Ι	Bacterial Genetics	2
	23-MB-242	II	Air, Water and Soil Microbiology	2
	23-MB-243	III	Practical Course based on theory papers 23-MB-241 and 23-MB-242	2

#### **Semester III**

#### 23-MB-231: Medical Microbiology and Immunology

[2 Credits; 36 Lectures]

#### [1 credit=15 hrs x 60 mins 900mins/50mins= 18 lectures]

#### **Course Outcomes:**

Students will be able to

CO1: Describe anatomy and physiology of human systems with associated pathogens.

CO2: Explain the concept of epidemiology of infectious diseases

CO3: Explain concept of chemotherapy, antibiotics, antagonism and synergism in drug

administration, antibiotic sensitivity and route of drug administration

CO4: Explain the process of Hematopoiesis, innate immunity and adaptive immunity, concept underlyingAntigens and Antibodies.

CO5: Relate genetics, biochemistry and inheritance of ABO and Rh blood group systems, medico legal applications of blood groups.

CO6: Differentiate between active and passive immunization, explain immunization schedule in India and the concept of immunization with examples of types of vaccines.

Unit I	Medical Microbiology	18
	Introduction to infectious diseases of following-	
	(Overview of pathogens associated with different systems and	
	common symptoms)	
	a) Respiratory System	
1	b) Gastrointestinal System and Liver	8
	c) Urinary System	
	d) Central Nervous System	
	e) Skin	

	1. Introduction to Epidemiology -	
	a) Definition of epidemiology and scope	
	b) Definition of epidemic, endemic and pandemic diseases	
2	c) Modes of transmission of infection	5
	d) Sources and reservoirs of infection	
	e) Disease prevention and control measures	
	Introduction to Chemotherapy	
	a) Definition of antibiotic, antiseptic, disinfectant drug	
	b) Characteristics of ideal chemotherapeutic agent	
3	c) Routes of drug administration	5
	d) Antagonism and synergism in drug administration	
	e) Antibiotic sensitivity testing	
Unit II	Immunology	(18)
	Immunity	(20)
1	Definition, Types (Innate and acquired, active and passive, humoraland	2
1	cell mediated)	2
	Formation of blood cells (Hematopoiesis)	
	a) Myeloid and lymphoid lineages and differentiation process	
2	b) Cells involved in innate immunity and adaptive immunity, their	5
	structure and function.	
	Antigens and antibodies: Definition and Concept	
3	a) Antigens: Examples of different antigens	2
5	b) Structure of antibody	2
	c) Introduction to cytokines and MHC molecules	
	a) ABO and Rh blood group systems	
	b) Bombay blood group	
4	c) Biochemistry of blood group substances	7
	d) Inheritance of ABH antigens - Problems based on ABO and Rh	
	blood group system	
	e) Medico legal applications of blood groups	

	Active and Passive Immunization	
	a) Active Immunization and Passive Immunization	
5	b) Types of vaccines: whole organism, inactivated, toxoid, combined, cellular fractions, recombinant and synthetic.	4
	c) Latest Immunization schedule in India	

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#### 23-MB-232: Bacterial Physiology and Fermentation Technology [2 Credits; 36 Lectures] [1 credit=15 hrs x 60 mins = 900mins/50mins= 18 lectures]

#### **Course Outcomes:**

Students will be able to -

CO1: Describe the components of holoenzyme, nomenclature and classification of enzymes, models of catalysis and effect of various parameters on enzymes.

CO2: Explain various glucose metabolic pathways with details such as structures and names of metabolites, names of enzymes and cofactors

CO3: Describe application of fermentation technology, screening, selection and maintenance of microbial

strains, design of fermentation media and fermenters, types of fermentations, working of fermenters,

consequences of contamination.

Unit I	Bacterial Physiology	(18)
	<b>Enzymes</b> i. Introduction to Enzymes: Properties of enzymes, Nature of active site, Structure ofactive site, commonly occurring amino acids at active site. Ribozymes, coenzymes, apoenzymes, prosthetic group and cofactors.	2
	ii. Nomenclature and classification as per IUB (up to class level).	2
1	<ul><li>iii. Models for catalysis–</li><li>a) Lock and key</li><li>b) Induced fit</li></ul>	1
	c) Transition state.	
	iv. Effect of pH and temperature, substrate concentration and enzyme concentration, activators and inhibitors of enzyme	3
	Bacterial Physiology	1
	i. Definitions of Metabolism, catabolism, anabolism, respiration and fermentation	
	ii. Metabolic pathways (with structures)	2
2	a) Embden-Meyerhof-Parnas pathway (Glycolysis). Entry of fructose and lactosein glycolysis. Substrate level phosphorylation.	2
	b) Hexose monophosphate pathway	1
	d) Phosphoketolase pathway (Hexose)	2
	e) TCA cycle (with emphasis on amphibolism) and Glyoxylate bypass f) Glycolysis and TCA cycle as central metabolic pathways.	

Unit II	Fermentation Technology	(18)
1	Concept of fermentation technology i. Microbial biomass- based fermentation (Biofertilizer, biopesticide and Probiotics) ii. Production of Primary metabolites (Organic acids, amino acids, vitamins and enzymes) iii. Production of Secondary metabolites (Antibiotics) iv. Production of recombinant products (insulin and growth hormones) v. Production of Fermented food products (Cheese, yoghurt) vi. Microbial biotransformation (Steroid transformation)	4
2	Strains of industrially important microorganisms: i. Desirable characteristics of industrial strain ii. Principles and methods of primary and secondary screening iii. Master, working and seed culture; development of inoculum iv. Preservation and maintenance of industrial strains.	5
3	<b>Design of a Fermenter (typical CSTR Continuous stirred</b> <b>Tank Reactor):</b> Different parts and their working	2
4	Monitoring of different fermentation parameters (Temperature, pH, aeration, agitation, foam)	2
5	Types of fermentations: Batch, continuous and dual	2
6	Media for industrial fermentations: Constituents of media (Carbon source, nitrogen source, amino acids, vitamins, minerals, water, buffers, antifoam agents, precursors, inhibitors and inducers)	2
7	Contamination: Sources, precautions and consequences	1

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#### 23-MB-233: Practical Course based on

#### 23-MB-231: Diagnostic Microbiology and Immunology

#### and

23-MB-232: Bacterial Physiology and Fermentation Technology

#### [2 Credits: 78 Lectures]

#### [1 credit=15hrs x 130 mins = 1950 mins/50 mins=39 lectures]

78 L distributed as 60 L for performing practicals and 18 L for internal evaluation

### **12 Practicals x 5 lectures = 60 Lectures**

#### **COURSE OUTCOMES:**

Students will be able to -

CO1: Measure cell dimensions by micrometry

CO2: Identify the blood group of blood sample

CO3: Demonstrate screening of organic acid/ antibiotic and amylase producing microorganisms...

CO4: Enrich and isolate Azotobacter and Rhizobium or Cyanobacteria and prepare biofertilizer

Expt.	Topics	No. of
No.		Practicals
1	Measurements of cell dimension by micrometry using 10x, 45x and 100x Objectives	1
2	Blood grouping: ABO, Rh and Bombay blood group (anti H Lectin test)	1
3	Cultural and Biochemical characteristization of bacteria Gram staining & motility Sugar utilization test, Sugar fermentation test, Enzyme detection – Gelatinase, Catalase, Oxidase	4
4	Staining techniques: i. Endospore staining         ii. Metachromatic granules	1
5	Primary screening of industrially important organisms:Screening and isolation of antibiotic and organic acid producing organism from soil by Crowded plate and Giant colony method	2
	Microorganisms producing industrially important enzyme-amylase	1
6	Enrichment, Isolation, Preparation and Application of Bioinoculants a) <i>Azotobacter</i> species and b) <i>Rhizobium</i> species	2
	Total	12

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## S. Y. B. Sc. Microbiology Syllabus (Semester IV) 23-MB-241: Bacterial Genetics [2 Credits; 36 Lectures] [1 credit=15 hrs x 60 mins = 900mins/50mins= 18 lectures]

#### **Course Outcomes:**

Students will be able to

CO1: Explain how the nature of genetic material was discovered and comprehend the structure of Nucleic acids

CO2: Comprehend the modes, rules and steps of DNA replication

CO3: Explain various types of mutations, types of mutagenic agents and their mechanism of action

CO4: Get an overview of gene expression and plasmid genetics

Unit I	Topics	(18)
	Understanding DNA:	7
1	i. Experimental evidence for nucleic acid as genetic material.	
	a. Discovery of transforming material (hereditary material):	
	b. Griffith's experiment	
	c. Avery and MacLeod experiment	
	d. Gierer and Schramm	
	e. Fraenkel-Conrat and Singer experiment (TMV virus)	
	f. Hershey and Chase experiment	
	ii. Types of nucleic acids (DNA and RNAs)	1
	iii. Structure of DNA	
	a. Structure of Nitrogen bases, Nucleoside, Nucleotide and	
	polynucleotide chain	2
	b. Bonds involved in DNA structure	_
	c. Different forms of DNA	
	iv. Prokaryotic DNA replication	
2	a. Models of DNA replication (Conservative, semi-	8

	conservative and Dispersive)	
	b. Meselson and Stahl's experiment (semi-conservative)	
	c. Basic mechanism of DNA replication	
	d. Enzymes, proteins and other factors involved in DNA	
	replication.	
	e. Modes of DNA replication Rolling circle mechanism, theta	
	and linear DNA replication	
Unit II	Topics	(18)
	i. Gene expression	4
	a. Concept of Genetic code and its properties	
1	b. Concept of transcription and translation	
	c. Levels of genome organization in prokaryotes	
	d. Levels of genome organization in eukaryotes	
	ii. Mutations and reversions	0
	Concept of Mutation and Types of mutations: Nonsense, Missense, Silent, Conditional lethal-temperature sensitive, Amber, Reverse, suppressor	,
	a. Spontaneous Mutation	
	Mechanism of spontaneous mutation	
	b. Concept of Induced Mutations	
2	<ul> <li>Base pair substitution (Transitions, Transversions), Insertions and deletions-Frame / Phase shift mutations</li> <li>Physical Mutagenic agent: UV and X-ray</li> </ul>	
2	Chemical mutagenic agents	
	• Base analogues (2 amino purine, 5 bromouracil) –Keto and Enol forms of Nitrogen bases.	
	HNO <sub>2</sub> , Alkylating agents	
	• Intercalating agents (EtBr, acridine orange)	
	Plasmid genetics	5
	a. Types and properties of plasmids.	
3	b. Concept of plasmid incompatibility, plasmid curing and amplification.	
	c. Plasmid replication Importance of plasmids in recombinant DNA Technology and other fields.	

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#### S. Y. B. Sc. Microbiology Syllabus (Semester IV)

#### 23-MB-242: Air, Water and Soil Microbiology

### [2 Credits; 36 Lectures]

#### [1 credit=15 hrs x 60 mins = 900mins/50mins= 18 lectures]

#### **Course Outcomes:**

CO1: The course will help them to get knowledge of the Air Microbiology, methods of air sampling, different types of air samplers, air sanitation and airborne infections.CO2: Deals with water microbiology including bacteriological analysis of water, methods of waterpurification, water borne infections and bacteriological standards of water quality.CO3: Understand Soil Microbiology, rhizosphere, composting and humus formation, biofertilizers, biocontrol agents and microbial interactions.

CO4: Acquire knowledge of carbon and nitrogen cycles with role of microorganisms.

Unit I	Air Microbiology and Water Microbiology	18
	i. Air Microbiology	
	a. Air flora	
	• Transient nature of air flora	
	• Droplet, droplet nuclei and aerosols	
	b. Methods of Air sampling and types of air samplers	
	• Impaction on solids	
	• Impingement in liquid	
	• Sedimentation	
	• Centrifugation	3
1	c. Air sanitation: Physical and chemical methods	2
	d. Airborne infections	1
	ii. Water Microbiology	
	a. Types of water: surface, ground, stored, distilled, mineral andde- mineralized water	1

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	b. Recommended Bacteriological standards of Water Quality	
	Maharashtra Pollution Control Board	
2	(MPCB) Main Functions of MPCB	
	Water quality standards for best designated usages	1
	Central Pollution Control Board	
	(CPCB) Main Functions of CPCB	
	Designated Best Use Water Quality Criteria	
	c. Water purification methods	2
	d. Water borne Infections	1
	e. Indicators of faecal pollution:	2
	Escherichia coli, Bifidobacterium, Streptococcus faecalis,	
	Clostridium perfringens,	
	New indicators: Campylobacter and Pseudomonas	
	f. Bacteriological analysis of water for potability	
	i. Bacteriological standards of potable water: Bureau of	
	Indian standards (BIS)	
	ii. World Health Organization (WHO)	
	iii. Presumptive coliform count	4
	iv. Confirmed test	
	v. Completed test	
	vi. Eijkman test	
	vii. Membrane filter technique	
	Soil Microbiology	18
	a. Rhizosphere microflora and its role in the rhizosphere	1
	b. Role of microorganisms in composting and humus formation	2
	c. Biofertilizers: Bacterial, Cyanobacterial, fungal and their large-	
	scale production	3
	d. Biocontrol agents: Bacterial, Viral, Fungal and their large-	
Unit	scale production	3
II	e. Brief account of microbial interactions:	
	Symbiosis, Neutralism, Commensalism, Competition, Ammensalism,	_
	Synergism, Parasitism and Predation	5

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f. Role of microorganisms in elemental cycles in nature: Carbon, Nitrogen

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#### S.Y. B. Sc. Microbiology Syllabus(Semester IV)

#### 23-MB-243: Practical Course based on 23-MB-241: Bacterial Genetics and MB-242: Air, Water and Soil Microbiology

[2 Credits: 78 Lectures]

[1 credit=15hrs x 130 mins = 1950 mins/50 mins=39 lectures]

### 78 L distributed as 60 L for performing practicals and 18 L for internal evaluation

**12 Practicals x 5 lectures = 60 Lectures** 

#### **Course Outcomes:**

Students will be able to

CO1: Estimate the diversity of microorganism by statistical analysis

CO2: Determine potability of drinking water using MPN test and membrane filtration

technique.

CO3: Isolate and identify pathogens E. coli, Staphylococcus aureus and Candida

from clinicalsample and characterize them by Gram staining, motility, cultural and

biochemical tests Demonstrate the use of physical and chemical mutagen to isolate

mutants.

Expt.No	Topics	No. of Practicals
1	Air Flora:	_
	<ul> <li>a. Diversity determination Simpson index.</li> <li>b. Determination of addimentation rate</li> </ul>	2
	b. Determination of sedimentation rate	
2	<ul> <li>a. MPN, Confirmed and Completed test.</li> <li>b. Membrane filter technique (Demonstration)</li> <li>c. Identification of <i>E. coli</i> from water sample as fecal indicator</li> </ul>	3
3	Tests for Biochemical characterization of bacteria i. Triple Sugar iron agar ii. IMViC test	4

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	<ul> <li>iii. Oxidative-fermentative test [Baird Parker's modification of Hugh and Leifson's oxidative- fermentative (OF) basal medium for Gram Positive and Hugh and Leifson's oxidative- fermentative (OF) basal medium for Gram negative; Public Health England, 2019]</li> </ul>		
4	<ul> <li>i. UV- survival curve</li> <li>ii. Induction of mutation by using physical mutagen (e.g. U V rays)</li> <li>iii. Isolation of auxotrophic mutants by Replica Plate</li> </ul>	2	
5	i. Visit to Industry/ Drinking Water treatment plant	1	
	Total	12	

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